

# Indoor Chemistry and Health - A Workshop Summary

Charles J. Weschler, J. R. Wells, Dustin Poppendieck, Heidi Hubbard, and Terri A. Pearce doi:10.1289/ehp.8271 (available at http://dx.doi.org/) Online 3 November 2005



**Indoor Chemistry and Health - A Workshop Summary** 

Charles J. Weschler<sup>1, 2</sup>

J. R. Wells<sup>3</sup>

Dustin Poppendieck<sup>4</sup>

Heidi Hubbard<sup>5</sup>

Terri A. Pearce<sup>3</sup>

1. International Center for Indoor Environment and Energy, Technical University of

Denmark, Lyngby, Denmark

2. Environmental and Occupational Health Sciences Institute, University of Medicine and

Dentistry of New Jersey & Rutgers University, Piscataway, NJ 08854

3. National Institute for Occupational Safety and Health, Morgantown, WV 26505

4. Environmental Resources Engineering, Humboldt State University, Arcata, CA 95521

5. Department of Civil Engineering, University of Texas at Austin, Austin, TX 78758

**Corresponding Author** 

Charles J. Weschler

Environmental and Occupational Health Sciences Institute

170 Frelinghuysen Road, Piscataway, NJ 08854

Email: weschlch@umdnj.edu

Phone: 732 235-4114; Fax: 732 530-1453

1

**Short Running Head:** Indoor Chemistry and Health

**Article Descriptor:** Risk Characterization

**Key Words:** Indoor pollutants, free radicals, hydroperoxides, biomarkers, inhalation exposure, asthma, allergies, environmental cancer, lung damage

**Acknowledgements:** We gratefully acknowledge the workshop participants who made presentations: John Balmes (University of California, San Francisco and University of California, Berkeley), William Cain (University of California, San Diego), Richard Corsi (University of Texas, Austin), Barbara Finlayson-Pitts (University of California, Irvine), Ann-Therese Karlberg (University of Gothenburg, Sweden), Howard Kipen (University of Medicine and Dentistry of New Jersey), Jane Koenig (University of Washington), Debra Laskin (University of Medicine and Dentistry of New Jersey), Claudia Miller (University of Houston), William Nazaroff (University of California, Berkeley), James N. Pitts, Jr. (University of California, Irvine), Tunga Salthammer (Wilhelm-Klauditz Institute, Germany), Dennis Shusterman (University of Washington), Jan Sundell (Technical University of Denmark), Charles J. Weschler (University of Medicine and Dentistry of NJ and Technical University of Denmark), Peder Wolkoff (National Institute of Occupational Health, Denmark), Paul Ziemann (University of California, Riverside). The workshop was funded by the National Institute for Occupational Safety and Health. We thank the Harvard School of Public Health for supporting the breaks at the workshop.

# **Abbreviations**

BF Blink Frequency

DALYs Disability Adjusted Life Years

ENO Exhaled Nitric Oxide

IE Indoor Environment

NIOSH National Institute for Occupational Safety and Health

NORA National Occupational Research Agenda

PCP Pentachlorophenol

RNI Reactive Nitrogen Intermediates

ROI Reactive Oxygen Intermediates

ROS Reactive Oxygen Species

TVOCs Total Volatile Organic Compounds

# **Manuscript Outline**

- I. Abstract
- II. Introduction
- III. Issues
  - i. Chemical reactions among indoor pollutants
- ii. Potential health effects associated with inhalation exposure to the products of indoor chemistry
  - iii. Techniques to study potential health effects
- IV. Recommendations
  - i. Priority research needs
  - ii. Testable hypotheses
- V. Conclusions
- VI. Disclaimer
- VII. References

#### Abstract

Chemicals present in indoor air can react with one another, either in the gas phase or on surfaces, altering the concentrations of both reactants and products. Such chemistry is often the major source of free radicals and other short-lived, reactive species in indoor environments. To what extent do the products of indoor chemistry affect human health? To address this question the National Institute for Occupational Safety and Health sponsored a workshop from 12 to 15 July 2004 at the University of California, Santa Cruz. Approximately 70 experts from eight countries participated. Objectives included enhancing communications between researchers in indoor chemistry and health professionals, as well as defining a list of priority research needs related to the topic of the workshop. The ultimate challenges in this emerging field are defining exposures to the products of indoor chemistry and developing an understanding of the links between these exposures and various health outcomes. The workshop on *Indoor Chemistry and Health* was a step towards meeting these challenges. This summary presents the issues discussed at the workshop and the priority research needs identified by the attendees.

#### Introduction

The National Institute for Occupational Safety and Health (NIOSH) established the National Occupational Research Agenda (NORA) in 1996 with input from more than 500 organizations and individuals. The indoor environment (IE) was included in the 21 NORA priority areas and the NORA IE Team was established. Its goals included focusing and facilitating research that would improve the health of workers in indoor environments.

As a means to address causes and prevention of specific building-related health effects, the NORA IE team conceived and sponsored a workshop entitled *Indoor Chemistry and Health*. "Indoor chemistry" is defined as reactions involving indoor pollutants, occurring either in the gas phase or on surfaces. In the absence of combustion, such chemistry is often the major source of free radicals and other short-lived reactive species in indoor environments. Approximately 70 scientists from 8 countries participated in this workshop held at the University of California, Santa Cruz from July 12-15, 2004. Disciplines represented included: atmospheric chemistry, chemical engineering, toxicology, medicine, epidemiology, architecture and public health. (A full participant list can be viewed at the NIOSH NORA IE web site: http://www2a.cdc.gov/nora/NTeamAct-IE-participants.htm).

A major goal was to promote communication between persons examining the health effects resulting from exposures to airborne pollutants and those studying outdoor and indoor chemistry. Experts from these respective disciplines made presentations, each of which was followed by group discussion. At the end of the workshop the participants

were charged with developing a list of research priorities and testable hypotheses at the interface between indoor chemistry and human health.

#### **Issues**

Presentations and discussions focused on three broad issues: 1) Chemical reactions among indoor pollutants; 2) Potential health effects associated with inhalation exposure to the products of indoor chemistry; and 3) Techniques to study potential health effects. Much of what follows comes from the presentations, for which we gratefully credit the presenters listed in the acknowledgements.

Chemical reactions among indoor pollutants. Dominant indoor processes include: i) oxidation reactions involving oxygen, ozone, hydroxyl (OH) and nitrate radicals (NO<sub>3</sub>); ii) acid/base reactions; iii) hydrolysis reactions and iv) decomposition reactions, often promoted by ultraviolet light and/or heat. Hydrolysis reactions are relatively slow and occur primarily on surfaces. The other processes can occur both in the gas-phase and on surfaces. Characteristic times associated with air-exchange, advective transport, diffusive transport, and first-order kinetics are important for indoor-pollutant dynamics and affect the outcome of reactions.

Ozone (O<sub>3</sub>) drives most indoor oxidative chemistry and can react at meaningful rates in the gas phase with nitric oxide (NO), nitrogen dioxide (NO<sub>2</sub>), and unsaturated organic compounds (e.g., terpenoids, sesquiterpenes, unsaturated fatty acids) to yield excited intermediates, hydroxyl and nitrate radicals, and oxygenated organic compounds

(Weschler 2004). The use of cleaning products containing both terpenes and glycol ethers in the presence of ozone can lead to oxidation of the glycol ethers via OH and perhaps NO<sub>3</sub> (Nazaroff and Weschler 2004); resultant products may include potentially allergenic peroxides and hydroperoxides (Karlberg et al. 2003). Modeling indicates that when ozone and NO<sub>2</sub> are present simultaneously, indoor NO<sub>3</sub> may be the dominant indoor oxidant; NO<sub>3</sub> levels as low as 1 ppt can compete effectively with O<sub>3</sub> and OH in oxidizing various terpenoids (Nazaroff and Weschler, 2004). There is a need for new analytical techniques to measure the products of indoor chemistry that are short-lived, highly reactive, thermally labile or highly oxidized – "stealth" chemicals. Oxidative chemistry has likely increased indoors over the past half-century, given increasing outdoor ozone levels, the greater indoor use of terpenoids (as odorants and cleaning products), and decreased ventilation rates.

Surface-to-volume ratios are much larger indoors than outdoors (roughly 3 versus 0.01 m<sup>2</sup>m<sup>-3</sup>), and, consequently, surface reactions tend to be more important indoors than out. At the molecular level, the fundamental principles of surface chemistry are the same outdoors, indoors, and within the respiratory tract. Indoor surfaces are diverse, including building materials, wall cavities, ducts, skin, clothing, dust, and airborne particles. As a consequence of surface chemistry, primary species can be altered/sorbed thereby influencing the amounts available for inhalation; many of the secondary species would not be present if indoor chemistry did not occur (e.g., products of ozone/carpet interactions (Weschler et al. 1992; Morrison and Nazaroff 2002)). Surface interactions influence subsequent human inhalation exposures to the constituents of environmental

tobacco smoke (Nazaroff and Singer, 2004); for example, acid-base chemistry influences nicotine's desorption from surfaces (Destaillats et al. 2005). In the case of carpet emissions, the presence of ozone influences aldehyde emissions with concentrations of some emitted oxidation products exceeding their odor thresholds (Morrison and Nazaroff 2002). As a consequence of sorption and re-emission from indoor surfaces, certain pesticides and fumigants that are transported indoors can remain at elevated concentrations and/or chemically transform for days or weeks. Malathion, a pesticide judged to be safe for humans, can be oxidized to malaoxon, a compound known to be toxic (Brown et al. 1993).

Other issues related to chemical reactions on surfaces include the interplay between sorption and surface reactions, the potential influence of surface chemistry on air quality in damp buildings and the aging/"regeneration" of surfaces. One of the first examples of indoor surface chemistry was the NO<sub>2</sub>/surface formation of HONO and HNO<sub>3</sub> (Pitts et al. 1985). It is now known that the resulting nitric acid on surfaces exists as an HNO<sub>3</sub>-H<sub>2</sub>O complex (Dubowski et al. 2004), yielding possible acidic, oxidizing, and nitrating surface films on interior walls. Air-water interfaces are a common feature of indoor environments and evidence indicates that chemistry is enhanced at such interfaces. Recent molecular dynamic simulations indicate that OH can be concentrated by a factor of six and O<sub>3</sub> by a factor of ten at such interfaces (Roeselova 2004). Similar behavior has been observed for some organic species (e.g., naphthalene). Indeed, surface chemistry within buildings may be dominated by interface reactions.

Building materials emit a myriad of reactive constituents and secondary products (derived from initial constituents). These include terpenoids, aliphatic aldehydes, phthalates, phenol, mono- and dicarboxylic acids, diisocyanates and various photoinitiators (Salthammer et al. 2002). An example of secondary emissions occurs in houses constructed with wooden studs treated with pentachlorophenol (PCP). Over time, PCP is transformed to tetrachloroanisole, giving occupants a highly undesirable odor (Gunschera et al. 2004). So-called "ecological" or "green" products are not necessarily free from adverse health effects; certain constituents such as terpenoids and linseed oil may be more chemically reactive than those from non-ecological products. Secondary emissions from such products may pose a greater health risk than the constituents for which their precursors are substitutes.

Thermal-desorption particle-beam mass-spectrometry has identified some of the more reactive products resulting from reactions of O<sub>3</sub> and NO<sub>3</sub> radicals with linear and cyclic alkenes (Ziemann 2002; 2003). Many of these products are relatively unstable and would not have been detected using conventional gas chromatographic/mass spectrometric (GC/MS) methods. Alcohols, carbonyls and carboxylic acids enhance the formation of secondary ozonides, as well as alkoxy and acyloxy hydroperoxides, from stabilized Criegee intermediates formed in O<sub>3</sub>/alkene reactions (Docherty et al. 2004). In other reaction pathways, carbonyls and carboxylic acids promote peroxyhemiacetal and polymer formation. Exploration of nitrate radical/alkene reactions has revealed that many products are multifunctional nitroxy, carbonyl, hydroxyl and hydroperoxyl

compounds. Some of the oxidation reaction products have vapor pressures low enough to lead to increased particle formation via molecular condensation (Ziemann 2002).

There are numerous gaps in our knowledge concerning indoor reactants and their products. A current need is measurements of the concentrations of OH, NO<sub>3</sub>, HO<sub>2</sub>, and CH<sub>3</sub>O<sub>2</sub> radicals under different indoor conditions for better understanding of their indoor chemistry (Sarwar et al. 2004). Indoor chlorine (Cl<sub>2</sub>) and chlorine oxide (HOCl, ClO<sub>2</sub>) chemistry has not received much attention; emission sources for such compounds include treated tap water, bleach and other cleaners. Anecdotal evidence exists for reactions between ClO<sub>2</sub> from tap water and new carpet leading to unpleasant odors. Chemical transformations occurring within heating, ventilating and air conditioning systems or in the immediate vicinity of the breathing zone ("near head chemistry") are potentially important, but have been little explored. Over time, additives in consumer products undergo chemical transformations (e.g., diphthalate esters hydrolyzing to alcohols and monoesters). However, the health consequences of exposure to such transformation products are largely unknown. Additionally, the ongoing introduction of new compounds into the indoor environment necessitates continual study of indoor emissions.

**Potential health effects associated with inhalation exposure to the products of indoor chemistry.** Organic compounds routinely measured in indoor air only partially, if at all, explain irritation complaints by building occupants (Wolkoff and Nielsen 2001). There needs to be a shift from what scientists can readily measure to what truly needs to be measured to improve exposure assessments, evaluations of health impacts, and

regulations. "Stealth chemicals" (see above) derived from indoor chemistry, may be partly responsible for sensory effects (Weschler and Shields 1997; Wolkoff and Nielsen 2001). Epidemiological studies support this hypothesis (Bluyssen et al. 1996; Sundell et al. 1993). For example, Sundell et al., in a study of 86 rooms in 29 office buildings, found that "lost TVOC" (lower TVOC concentrations in the room air than in the supply air) was inversely proportional to sick-building-syndrome symptoms. The strong association between "lost TVOC" and occupant symptoms provided some of the earliest evidence for an association between chemical transformations of indoor pollutants and adverse health effects.

Human sensitivity to complex mixtures of short- and long-lived radicals, ozonides, organic acids, and other oxygenated intermediates species remains unknown. Using a mouse bioassay, researchers have demonstrated that terpene oxidation products -- in the ozone/R-limonene, ozone/α-pinene and ozone/isoprene systems -- are more irritating to the upper airways than terpenes or ozone alone. (Wolkoff et al. 1999; Rohr et al. 2002). The currently identified oxidation products are insufficient to fully explain the irritation response and unidentified oxidation products could be contributing to the effects. Short-lived species may be responsible, since the bioresponse was diminished in experiments conducted at higher relative humidity and with longer reaction times (Wilkins et al, 2003).

Human subjects (female, late 20's, no serious sensitivities) were exposed to a mixture of 40 ppb of O<sub>3</sub> and 23 VOCs, including two terpenes, the same mixture without O<sub>3</sub>, or air

with a lower concentration of the VOC mixture (Fiedler et al. 2002). Monitored responses were both psychological (symptoms, odor ratings) and physiological (lung function, neuroendocrine, neurobehavioral, and inflammatory markers). The mixture that included O<sub>3</sub> had significantly higher concentrations of formaldehyde, glyoxal, hydrogen peroxide and secondary organic aerosols. Nonetheless, participant responses were similar regardless of exposure condition. Hence, for the time-scale (~ 2 hrs) and sensitivity of these experiments, there was no pronounced association between exposure to the products of indoor chemistry and the effects monitored in this study.

Human eye exposures have been used as a tool for evaluating exposures to products of ozone/alkene chemistry. No change in blink frequency (BF) was observed for ozone or limonene alone or the ozone/isoprene mixture, but there was a significant increase in BF upon exposure to a mixture of ozone/limonene or ozone/NO<sub>2</sub> (Kleno and Wolkoff 2004). Increased relative humidity decreased BF. Additional factors to examine are the role of free radicals as well as fine and ultrafine particles in blinking and eye irritation; blink frequency response versus chemical product concentrations; the physiological mechanisms; and the nature of chemicals that disrupt the tear film. An overarching question is whether eye blink rates provide an early warning of a health effect.

The anatomy of the upper airway and its responses to irritants such as  $O_3$  and chlorine are relevant to potential health effects caused by products of indoor chemistry (Shusterman 2003; Shusterman and Avila 2003). Considering the airways as a collection and filtering system designed to condition air for use by the human body allows for discrete

compartmentalization based on function. The water solubility of a pollutant influences its impact on the airway – the most water soluble chemicals affect the eyes, nose and throat; less water soluble chemicals affect the middle airway (bronchial tubes); and the least water soluble chemicals affect the lower airway (deep lung and alveoli).

There is evidence that inhaled oxidant pollutants produce oxidative stress coupled with upregulation of inflammatory cytokine production in the airways of asthmatics. Genetic polymorphisms in key antioxidant enzymes may predict susceptibility to cytotoxic tissue injury from oxidative stress (Bergamaschi et al. 2001). Reactive oxygen species (ROS) found in or generated by diesel particles, fly-ash from oil furnaces, O<sub>3</sub>, and other oxidant air pollutants can damage lipids, proteins and DNA, and initiate a chain of events started by macrophages and targeting pollutant capture and neutralization (Arjomandi et al. 2005). Present knowledge indicates that: 1) pollutant-induced oxidative stress leads to pro-inflammatory gene expression through multiple pathways; 2) oxidant pollutants can enhance responses to environmental allergens; and 3) there are systemic effects of pollutant-induced oxidative stress in the lung that are important in cardiovascular toxicity.

Ozone provides a good example of the consequences of inhaling a reactive pollutant. The pulmonary effects include: airway hypersecretion, decreased lung function, epithelial cell damage and inflammation. Ozone exposure activates macrophages, the second most potent secretory cells in the body and critical mediators of inflammatory response.

Macrophage over-activation, with excessive production of cytotoxic and pro-

inflammatory mediators, can contribute to tissue injury. Mediators include cytokines, reactive oxygen intermediates (ROI) such as superoxide, hydrogen peroxide and hydroxyl radicals, and reactive nitrogen intermediates (RNI) such as nitric oxide and peroxynitrite (Laskin et al. 2004; Fakhrzadeh et al. 2004). Studies with ozone-exposed rats have shown that macrophages release TNFα and IL-18 leading, through a series of steps, to NO production and ultimately tissue injury. Blocking macrophage NO production by gadolinium chloride has been shown to prevent the observed ozone-induced tissue injury (Pendino et al. 1995), providing evidence for RNI's role in tissue injury. The extent to which inhaling other reactive species (e.g., peroxy radicals or hydroperoxides) results in over-activation of macrophages is not known.

Dermal exposures are also of concern. Karlberg et al. (1994) has shown that air-oxidation of limonene produces contact allergens. These include limoneneoxide, carvone and a series of hydroperoxide isomers. Similarly, the oxidation of linalool yields allergenic hydroperoxide isomers (Skold et al. 2002). Special methods are required to isolate and identify hydroperoxides, which are unstable and readily form the corresponding aldehyde. When glycol ethers (ethoxylated surfactants) are exposed to air, allergenic oxidation products are also formed, although not as quickly as with terpenoids (Karlberg et al. 2003). These air-oxidation reactions are normally slow. However, some allergenic oxidation products can be formed at much faster rates through ozone-initiated oxidation processes.

Although the workshop focused on potential acute effects that might results from exposure to the products of indoor chemistry, it was agreed that researchers should also be mindful of potential chronic effects, especially cancer.

**Techniques to study potential health effects.** There are multiple methods to study the impact of pollutants on the respiratory tract, including acoustic rhinometry, nasal peak inspiratory flow, nasal scraping, nasal lavage, olfactory testing and trigeminal nerve sensory acuity. Physiological changes such as watery eyes and nose, or changes in the cells lining the contact surfaces, can be indicators of irritation and may be quantifiable.

Biomarkers for exposure to selected products of indoor chemistry would be of obvious utility. Changes in exhaled nitric oxide (eNO) concentrations have been used to track asthma and have been associated with exposure to outdoor air pollution (Koenig et al. 2003). Nitric oxide is ubiquitous in the body and is elevated in exhaled breath of asthmatics or persons having an asthma attack. Increases in 2.5 micrometer particulate matter (PM<sub>2.5</sub>) and light-absorbing carbon particles have been associated with airway inflammation, measured as increases in eNO in older subjects with asthma (although a similar increase was not observed in older subjects with chronic obstructive pulmonary disease). Given that eNO is a marker of oxidative stress, exposures to certain products of indoor chemistry (e.g., hydroxyl radicals, nitrate radicals, ozonides and hydroperoxides) may also lead to increases in eNO. However, the rapid oxidation of NO by certain oxidants may complicate its utility as a biomarker.

Chemesthesis -- the "feel" of a chemical -- describes chemically provoked irritation. Only three receptors are involved in chemesthesis versus over three hundred for olfaction.

Odor perception tends to increase gradually with increasing chemical concentration, while chemesthesis requires a threshold concentration to elicit response and then increases fairly rapidly (Cometto-Muniz et al. 2005). Chemicals tend to stimulate at equal fractions of their saturation vapor pressure. For molecules above a certain size, subjects are not able to feel the chemical; the reason for this is not well understood. For non-reactive molecules, the chemesthesis threshold for a brief exposure is typically greater than 1 ppm; for reactive molecules it may be lower. In the case of a limonene/ozone mixture (at realistic concentrations), subjects' chemesthesis response increased over time. The duration of the exposure has an amplifying effect on both chemesthesis magnitude and sensitivity (Cometto-Muniz et al. 2004).

A subset of building occupants is especially susceptible to pollutant exposures (Miller 1997). Such individuals can serve to alert health professionals to problematic indoor environments, including those with elevated species derived from indoor chemical reactions. There was a brief discussion regarding methods to identify such individuals.

Workshop participants agreed that it is crucially important to understand exposures and that insufficient time had been spent discussing exposures of different populations to the products of indoor chemistry. Knowledge regarding actual exposures and intakes is extremely important in making eventual connections with health outcomes. This is an area requiring much more attention.

#### Recommendations

A common theme running through workshop discussions was the need to better characterize and understand the "reacting" indoor environment, with an emphasis on the chemicals that most affect human health - the "biologically relevant" compounds. New methods need to be developed that can detect some of the elusive, short-lived highly reactive products.

At the conclusion of the presentations, the participants were split into seven groups, each charged with developing a list of at least three research priorities and one or more hypotheses, which were subsequently discussed and prioritized by the full set of participants.

**Priority Research Needs.** The resulting list of research needs can be grouped into 6 categories (the first three were judged to be most important):

- 1. Exposure. Conduct targeted exposure studies for specific compounds formed by reactions among indoor pollutants, as well as reaction product precursors. Focus on health-relevant (acute and chronic) compounds. Incorporate methods demonstrated to be useful in studies of outdoor pollutants. Take advantage of existing exposure biomarkers (or identify new biomarkers) for targeted products of indoor chemistry.
- **2. Modeling/Measurements/Model Evaluation.** A) Evaluate indoor chemistry models by measuring the concentrations of key reaction byproducts (e.g., hydroxyl, nitrate, hydroperoxy and methylperoxy radicals) under a variety of indoor conditions. Employ

existing techniques that have been successfully applied to outdoor air. Such measurements would be used to evaluate and improve the models. The improved models, in turn, would be used to focus additional measurements. Ultimately, targeted measurements of key reaction products should occur. B) Develop integrated pharmacokinetic models addressing potential irritation, inflammation and allergic responses initiated by the reaction products judged to be the most biologically significant.

- 3. Risk Assessment. Evaluate the health risks posed by the known products of indoor chemistry. This could be done using "Disability Adjusted Life Years". (DALYs equal the sum of years of premature mortality plus years of illness or injury modified by appropriate weighting factors due to a particular disease or risk factor (Anand and Hanson 1997)). Further risk assessment of reaction products would be based on toxicology, structure activity relationships and epidemiologic studies addressing both cancer and non-cancer endpoints.
- **4. Tissue Irritation.** Evaluate the contribution of the products of indoor chemistry to irritation, especially mucosal irritation, and the susceptibilities of various target organs. Evaluate the consequences of chemical reactions that might occur on biological surfaces such as skin or human lung tissue.
- **5. Screening Test.** Develop a rapid screening test (for example, in-vitro cell bioassays) that would permit initial health-effects evaluation of compounds generated by reactions among indoor pollutants.
- **6. Integrated Program Addressing Inflammation, Allergies and Asthma.** Screen products of indoor chemistry for their potential to exacerbate allergies or asthma and irritate mucous membranes. Following screening, evaluate the public's exposures to the

compounds of greatest concern coupled with detailed evaluations of these compounds' toxicology.

**Testable Hypotheses.** The participants agreed that the subject of the workshop itself could be stated as a testable hypothesis – that products of indoor chemistry adversely affect human health. More specifically, the testable hypotheses offered by the participants covered four areas:

- **1. Mucosal Irritation.** Chemical transformations of indoor pollutants yield products that contribute to mucosal irritation and inflammation.
- **2. Allergies.** Selected products of indoor chemistry can promote allergies (type 1 hypersensitivity).
- **3. Intervention.** Removing ozone or sources of chemically reactive pollutants will lead to health improvements in environments where the intervention occurs (by limiting the products of ozone-initiated chemistry).
- **4. Ecological labels.** Chemical transformations of constituents found in various indoor "green" or "ecological" materials subsequently contribute to, rather than mitigate, health problems.

The focused research needs identified at the Indoor Chemistry and Health Workshop are consistent with the broader research needs identified in the 2002 NORA IE Team publication: "Improving the health of workers in indoor environments: priority research needs for a national occupational research agenda" (Mendell et al. 2002).

### **Conclusions**

In the developed world, human exposure to airborne chemicals is dominated by indoor exposures. Inhalation of airborne pollutants is known to adversely affect human health, producing both acute and chronic effects. These include mucous membrane irritation, allergies and asthma, cardiopulmonary effects and cancer. Some of the species inhaled indoors come from outdoors; some come directly from materials and products used indoors, and some are a consequence of chemical reactions occurring in the indoor environment. Certain chemical processes are continually occurring indoors (e.g., hydrolysis of esters on indoor surfaces). Other chemical processes are occurring intermittently, varying with time of day, day of week, season and location (e.g., ozoneinitiated oxidation of terpenoids). Discussions throughout the workshop made it clear that research designed to evaluate the potential impacts that such indoor chemical processes have on human health has only just begun. The challenges in this emerging field are to define exposures to products of indoor chemistry and develop an understanding of the links between these exposures and various health outcomes. For this to happen, health professionals and associated researchers must be aware of indoor chemistry. The workshop on Indoor Chemistry and Health was an early step in developing such awareness.

#### Disclaimer

The findings and conclusions in this paper are those of the authors and do not necessarily represent the views of the National Institute for Occupational Safety and Health.

## References

Anand S, Hanson K 1997. Disability-adjusted life years: a critical review. Journal of Health Economics 16(6):685-702.

Arjomandi M, Witten A, Abbritti E, Reintjes K, Schmidlin I, Zhai W et al. 2005.

Repeated exposure to ozone increases alveolar macrophage recruitment into asthmatic airways. Am J Respir Crit Care Med 172:427-32.

Bergamaschi E, De Palma G, Mozzoni P, Vanni S, Vettori MV, Broeckaert F et al. 2001. Polymorphism of quinone-metabolizing enzymes and susceptibility to ozone-induced acute effects. Am J Respir Crit Care Med. 163(6):1426-31.

Bluyssen PM, Fernandes ED, Groes L, Clausen G, Fanger PO, Valbjorn O et al. 1996.

European indoor air quality audit project in 56 office buildings. Indoor Air 6:221-238.

Brown MA, Petreas MX, Okamoto HS, Mischke TM, Stephens RD 1993. Monitoring of malathion and its impurities and environmental transformation products on surfaces and in air following an aerial application. Environ Sci Technol 27:388-397.

CDC (Center for Disease Control). 2005. NIOSH NORA Indoor Environment Team Activities and Conferences. Attendees for Workshop. Available:

http://www2a.cdc.gov/nora/NTeamAct-IE-participants.htm [accessed 30 October 2005].

Cometto-Muniz JE, Cain WS, Abraham MH. 2004. Chemosensory additivity in trigeminal chemoreception as reflected by detection of mixtures. Exp Brain Res 158:196-206.

Cometto-Muniz JE, Cain WS, Abraham MH. 2005. Odor detection of single chemicals and binary mixtures. Behav Brain Res 156:115-123.

Destaillats H, Singer BC, Lee SK, Gundel LA. 2005. Acid-base interactions of nicotine and pyridine on surfaces. in Proceedings of Indoor Air 2005, Yang X, Zhao B, Zhao R (eds) Tsinghua University Press:1700-1704.

Docherty KS, Kumboonlert K, Lee IJ, Ziemann PJ. 2004. Gas chromatography of trimethylsilyl derivatives of alpha-methoxyalkyl hydroperoxides formed in alkene-O<sub>3</sub> reactions. J Chromatog A 1029:205-215.

Dubowski Y, Sumner AL, Menke EJ, Gaspar DJ, Newberg JT, Hoffman RC et al. 2004. Interactions of gaseous nitric acid with surfaces of environmental interest. Phys Chem Chem Physics 6(14):3879-3888.

Fakhrzadeh L, Laskin JD, Laskin DL. 2004. Ozone-induced production of nitric oxide and TNF-alpha and tissue injury are dependent on NF-kappa B p50. Am J Physiol Lung Cell Mol Physiol 287:L279-L285.

Fiedler N, Zhang J, Fan T, Kelly-McNeil K, Lioy P, Gardner C et al. 2002. Health effects of a volatile organic mixture with and without ozone. In: Indoor Air 2002: Proceedings of the 9th International Conference on Indoor Air Quality and Climate, Levin H (ed.), 2:596-601.

Gunschera J, Fuhrmann F, Salthammer T, Schulze A, Uhde E. 2004. Formation and emission of chloroanisoles as indoor pollutants. Env Sci Poll Rch 11:147-151.

Karlberg AT, Shao LP, Nilsson U, Gafvert E, Nilsson JLG. 1994. Hydroperoxides in oxidized d-limonene identified as potent contact allergens. Arch Dermatol Res 286:97-103.

Karlberg AT, Bodin A, Matura M. 2003. Allergenic activity of an air-oxidized ethoxylated surfactant. Contact Dermatitis 49:241-247.

Kleno J, Wolkoff P. 2004. Changes in eye blink frequency as a measure of trigeminal stimulation by exposure to limonene oxidation products, isoprene oxidation products and nitrate radicals. Int Arch Occup Environ Health 77:235-243.

Koenig JQ, Jansen K, Mar TF, Lumley T, Kaufman J, Trenga, CA et al. 2003.

Measurement of offline exhaled nitric oxide in a study of community exposure to air pollution. Environ Health Perspect 111: 625-1629.

Laskin DL, Heck DE, Laskin JD. 2004. Role of nitric oxide in acute lung injury. Shock 21:78-78.

Mendell MJ, Fisk WJ, Kreiss K, Levin H, Alexander D, Cain WS et al. 2002. Improving the health of workers in indoor environments: Priority research needs for a national occupational research agenda. Am J Public Health 92:1430-1440.

Miller CS. 1997. Toxicant-induced loss of tolerance--an emerging theory of disease? Environ Health Perspect 105(Suppl 2):445-453.

Morrison GC, Nazaroff WW. 2002. Ozone interactions with carpet: Secondary emissions of aldehydes. Environ Sci Technol 36:2185-2192.

Nazaroff WW, Singer BC. 2004. Inhalation of hazardous air pollutants from environmental tobacco smoke in US residences. J Expo Anal Environ Epidemiol 14:S71-S77.

Nazaroff WW, Weschler CJ. 2004. Cleaning products and air fresheners: exposure to primary and secondary pollutants. Atmos Environ 38:2841-2865.

Pendino KJ, Meidhof TM, Heck DE, Laskin JD, Laskin DL. 1995. Inhibition of macrophages with gadolinium chloride abrogates ozone-induced pulmonary injury and inflammatory mediator production. Am J Respir Cell Mol Biol 13:125-132.

Pitts Jr. JN, Wallington TJ, Biermann HW, Winer AM. 1985. Identification and measurement of nitrous acid in an indoor environment. Atmos Environ 19:763-767. Roeselová M, Vieceli J, Dang LX, Garrett BC, Tobias DJ. 2004. Hydroxyl radical at the air-water interface. J Am Chem Soc 126:16308-16309.

Rohr AC, Wilkins CK, Clausen PA et al. 2002. Upper airway and pulmonary effects of oxidation products of (+)-alpha-pinene, d-limonene, and isoprene in BALB/c mice. Inhal Toxicol 14:663-684.

Salthammer T, Bednarek A, Fuhrmann F et al. 2002. Formation of organic indoor air pollutants by UV-curing chemistry. J Photochem Photobiol A-Chem 152:1-9. Sarwar G, Olson DA, Corsi RL, Weschler CJ. 2004. Indoor fine particles: The role of terpene emissions from consumer products. J Air Waste Manag Association 54:367-377. Shusterman D. 2003. Toxicology of nasal irritants. Current Allergy And Asthma Reports 3:258-265.

Shusterman D, Avila PC. 2003. Real-time monitoring of nasal mucosal pH during carbon dioxide stimulation: Implications for stimulus dynamics. Chemical Senses 28:595-601. Skold M, Borje A, Matura M, Karlberg AT. 2002. Studies on the autoxidation and sensitizing capacity of the fragrance chemical linalool, identifying a linalool hydroperoxide. Contact Dermatitis 46:267-272.

Sundell J, Andersson B, Andersson K, Lindvall T. 1993. Volatile organic compounds in ventilating air in buildings at different sampling points in the buildings and their relationship with the prevalence of occupant symptoms. Indoor Air 3:82-93.

Weschler CJ, Hodgson AT, Wooley JD. 1992. Indoor chemistry: ozone, volatile organic compounds, and carpets, Environ Sci Technol 26:2371-2377.

Weschler CJ, Shields HC 1997. Potential reactions among indoor pollutants. Atmos Environ 31:3487-3495.

Weschler CJ. 2004. Chemical reactions among indoor pollutants: what we've learned in the new millennium. Indoor Air 14:184-194.

Wilkins CK, Wolkoff P, Clausen PA, Hammer M, Nielsen GD 2003. Upper airway irritation of terpene/ozone oxidation products (TOPS). Dependence on reaction time, relative humidity and initial ozone concentration. Toxicol Lett 143:109-114.

Wolkoff P, Clausen PA, Wilkins CK, Hougaard KS, Nielsen GD 1999. Formation of strong airway irritants in a model mixture of (+)-alpha-pinene/ozone. Atmos Environ 33:693-698.

Wolkoff P, Nielsen GD 2001. Organic compounds in indoor air – their relevance for perceived indoor air quality. Atmos Environ 35:4407-4417.

Ziemann, PJ 2002. Evidence for low-volatility diacyl peroxides as a nucleating agent and major component of aerosol formed from reactions of O-3 with cyclohexene and homologous compounds. J Phys Chem A 106:4390-4402.

Ziemann PJ 2003. Formation of alkoxyhydroperoxy aldehydes and cyclic peroxyhemiacetals from reactions of cyclic alkenes with  $O_3$  in the presence of alcohols. J Phys Chem A 107:2048-2060.